

The involvement of the Epidermal Growth Factor Receptor (EGFR) in the successful Growth Hormone (GH) signalling and the role of p21 in the negative regulation of the GH/GHR and EGF/EGFR pathways, in Growth Hormone Transduction Defect (GHTD).

Eirini Kostopoulou¹, Andrea Paola Rojas-Gil², Alexia Karvela¹, Bessie E. Spiliotis¹

1. Pediatric Endocrine Research Laboratory, Division of Pediatric Endocrinology and Diabetes, Department of Pediatrics, University of Patras School of Medicine, Patras, Greece. 2. Faculty of Human Movement and Quality of Life Sciences, Department of Nursing, University of Peloponnese.

Introduction

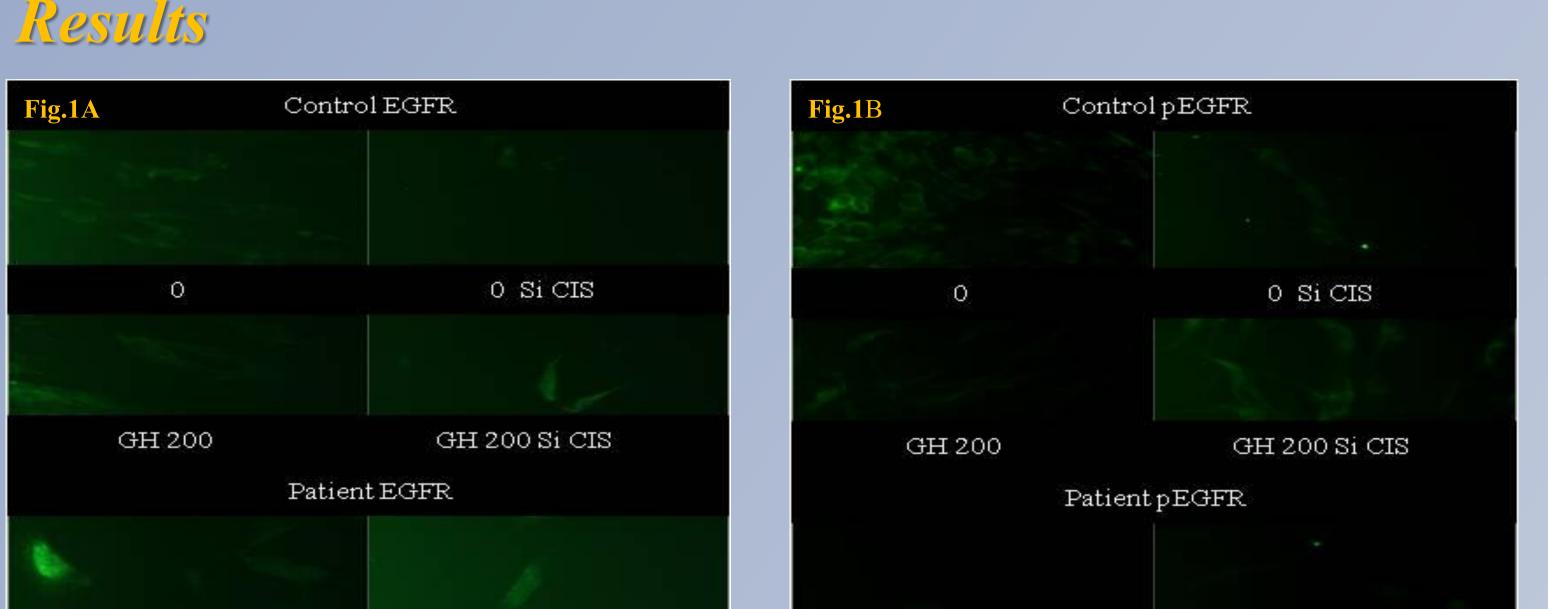
The authors have nothing to disclose

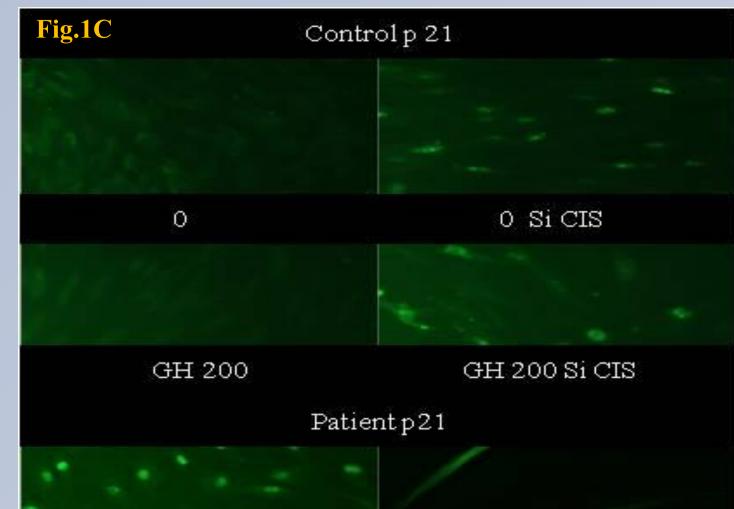
GHTD is characterized by:

✓ severe short stature and "catch-up" growth after hGH treatment.

✓ impaired STAT3 phosphorylation \checkmark overexpression of the E3 ubiquitin ligase, CIS

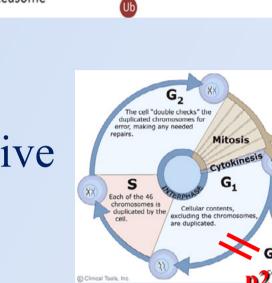
> H/GHR signalling athwavs





0 Si CIS

J GHR Cytosolic Proteasome Transcription of target genes Nucleus Growth and metabolism TRENDS in Genetics ✓ overexpression of the negative regulator of cell cycle, p21.



 $(\downarrow) (E1) (0) (E1) (0) (E1) (0) ($

by STAT3 phosphorylation is restored simultaneous induction of GHTD fibroblasts with 200µg/L GH and gene silencing of CIS with $1000 \mu g/L$ (GH200/siRNA) GH or (GH1000) alone.¹

Crosstalk between the GH and EGF pathways has been described in literature. ^{2,3,4}

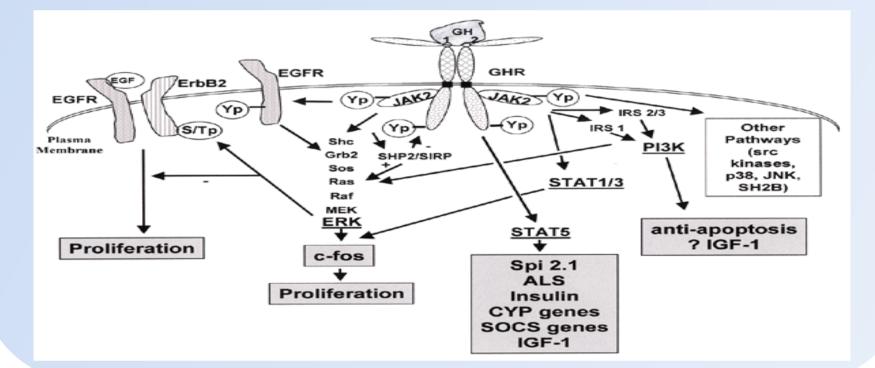
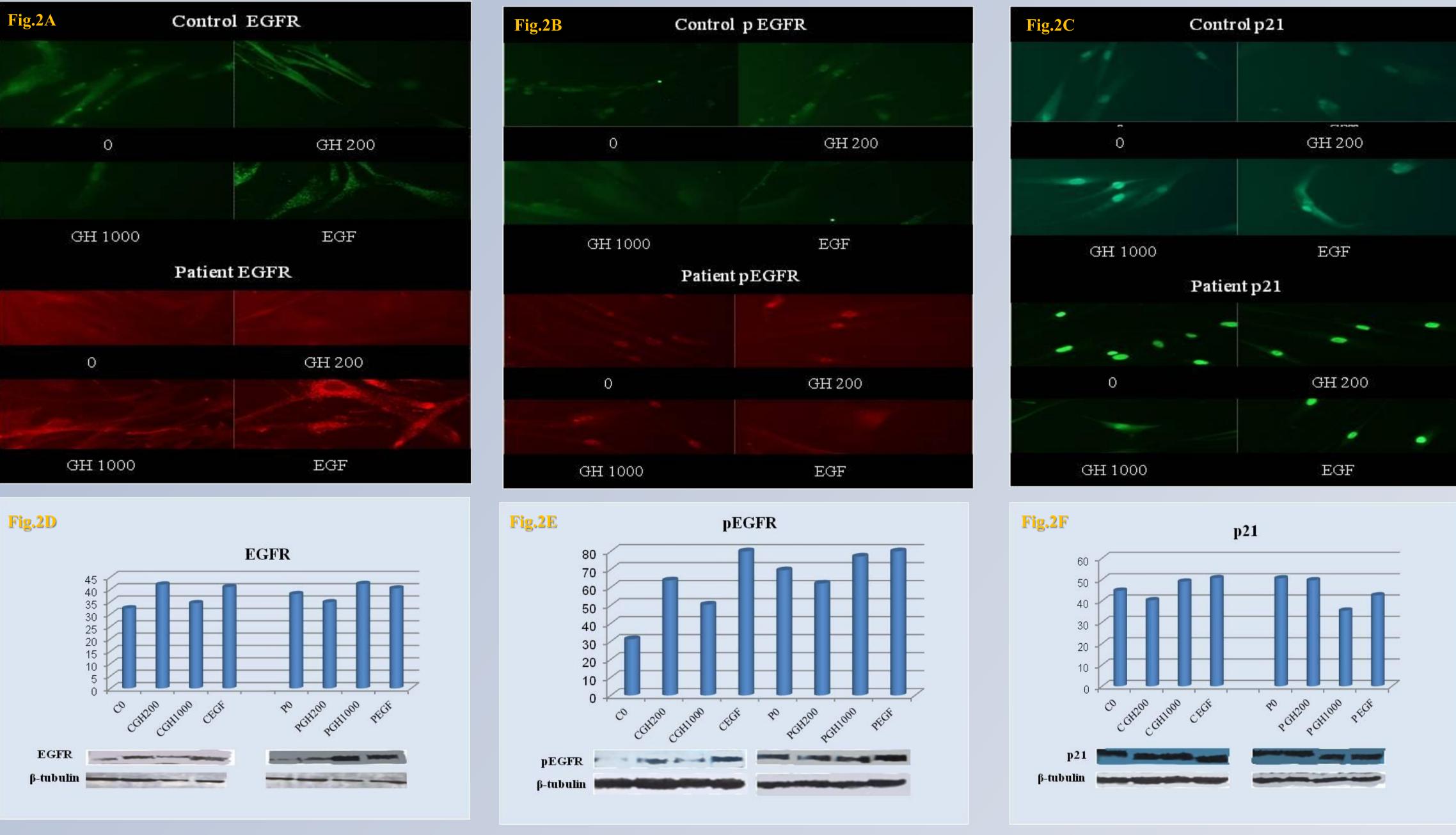
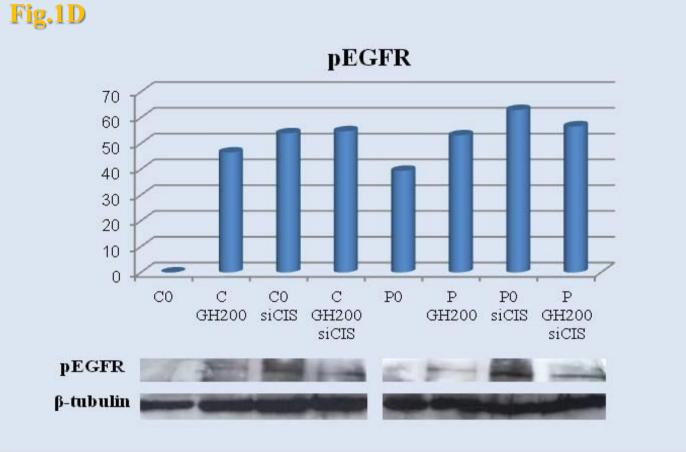


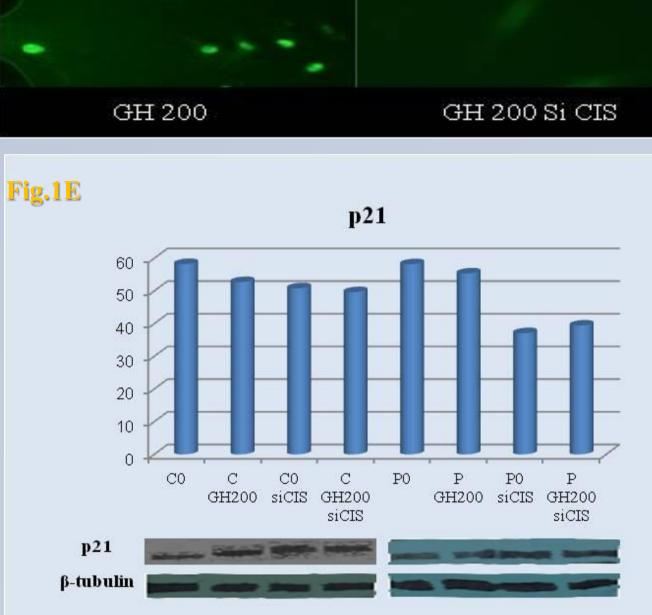


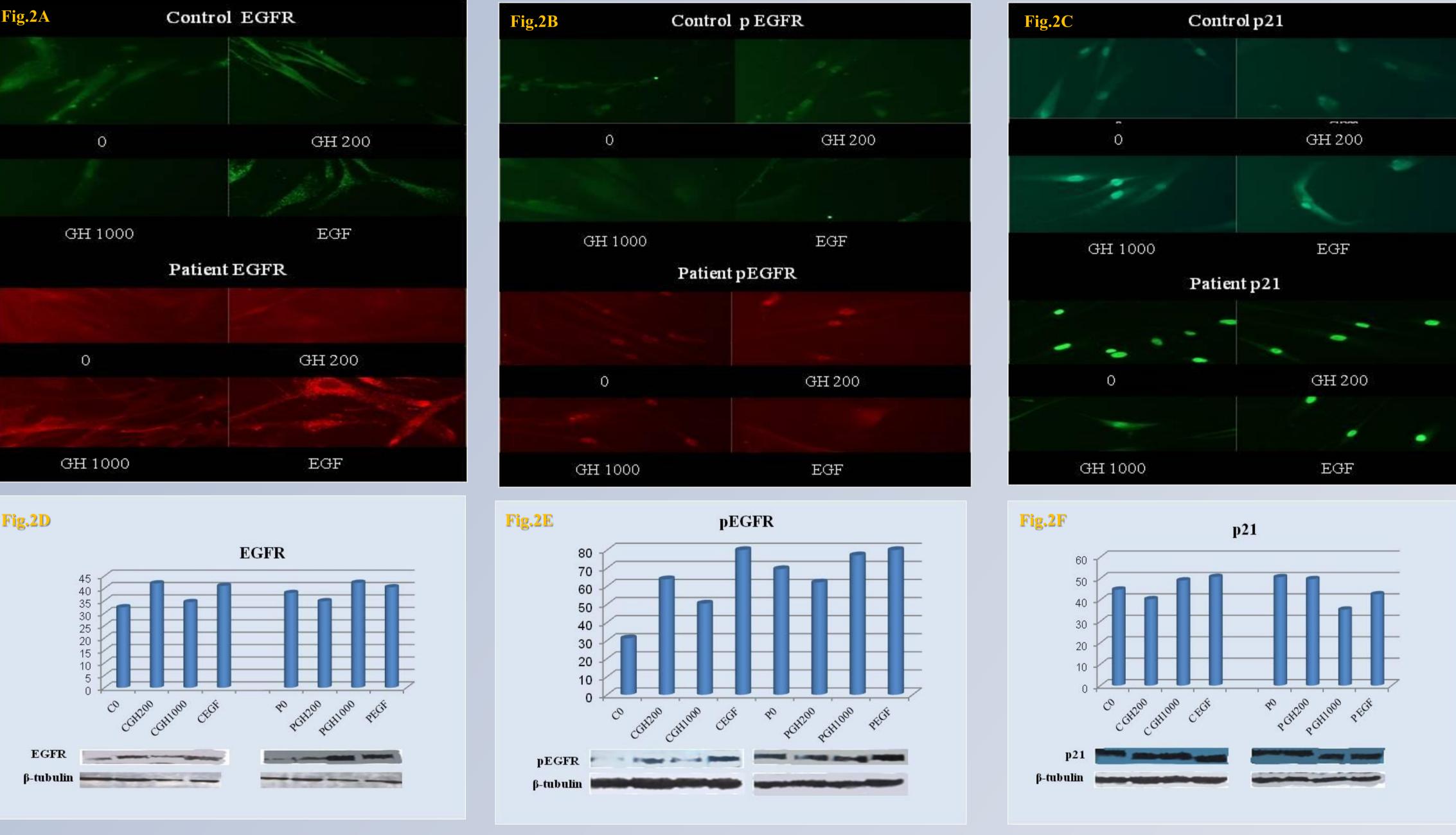
Figure 1. After GH200/siCIS: i) the protein expression and the membrane localization of EGFR (1A) and pEGFR (1B & 1D) were increased, especially in the patient, ii) the protein expression and the nuclear (anti-proliferative) localization of p21 were reduced in the patient (1C & 1E).

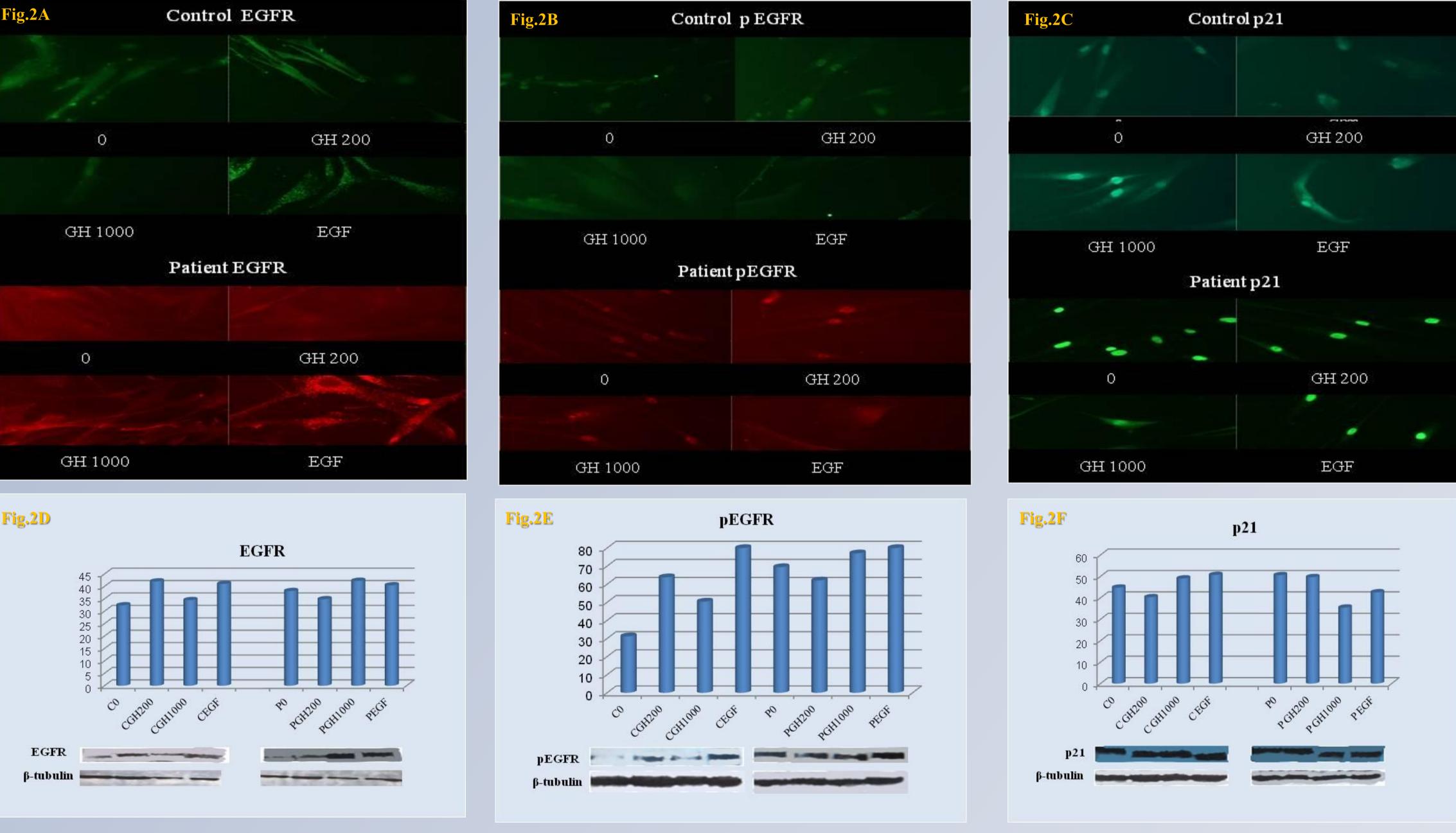












Objective

The involvement of the EGFR in successful GH signalling and the role of p21 in the negative regulation of the GH/GHR and EGF/EGFR pathways in GHTD.

Methods

Fibroblast cultures were developed from gingival biopsies of 1 GHTD patient and 1 control.

protein expression and The cellular localization of EGFR, pEGFR and p21 were studied by Western Immunoblotting and Immunofluorescence, respectively:

a) At the basal state and after induction with 200µg/L hGH (GH200), either with or without siRNA CIS.

Figure 2.

- + After the inductions where successful GH signalling was achieved (GH200 for the control and GH1000 for the patient):
- the protein expression and the membrane localization of EGFR (2A & 2D) and pEGFR (2B & 2E) were increased and
- <u>ii</u>. the protein expression and nuclear localization of p21 (2C & 2F) were reduced,
- ➡ After induction with EGF:
 - the protein and membrane expression of EGFR and pEGFR (2A, 2B, 2D & 2E) were increased similarly in the

b) At the basal state and after inductions with 200µg/L hGH (GH200), 1000µg/L hGH (GH1000) or 50 ng/ml EGF.

- control and in the patient and
- ii. The protein expression of p21 (2F) was increased in the control and reduced in the patient compared to the basal state.

Conclusions In the fibroblasts of the GHTD patients:

When CIS is reduced, either after silencing of the CIS gene or after a higher dose of hGH (GH1000), the EGFR is activated, nuclear p21 (anti-proliferative effect) is reduced and GH signalling is successful.

2) The EGFR is involved in successful GH signalling, more intensely than in the fibroblasts of the control child. The EGFR may be involved in the mechanism of "catchup" growth seen in the GHTD patients when exogenous hGH is administered.

3) p21 seems to participate also in negative regulation of the EGF/EGFR pathway in the control.

References

1) Rojas-Gil AP, Kostopoulou E, Karageorgou I, Kamzelas K & Spiliotis BE. Increased growth hormone receptor (GHR) degradation due to over-expression of cytokine inducible SH2 domain-containing protein (CIS) as a cause of GH transduction defect (GHTD), J Pediatr Endocr Met 2012; 25(9-10): 897–908.

2) Kim SO, Houtman JC, Jiang J, Ruppert JM, Bertics PJ & Frank SJ. Growth hormone-induced alteration in ErbB-2 phosphorylation status in 3T3–F442A fibroblasts. J Biol Chem 1999; 274:36015–36024.

3) Huang Y, Kim SO, Jiang J & Frank SJ. Growth hormone-induced phosphorylation of EGF receptor in 3T3–F442A cells: modulation of EGF-induced trafficking and signaling. J Biol Chem 2003; 278:18902– 18913.

4) Yamauchi T, Ueki K, Tobe K, Tamemoto H, Sekine N, Wada M, Honjo M, Takahashi M, Takahashi T, Hirai H et al. Tyrosine phosphorylation of the EGF receptor by the kinase Jak2 is induced by growth hormone. Nature 1997; 390:91–96.



DOI: 10.3252/pso.eu.54espe.2015

