

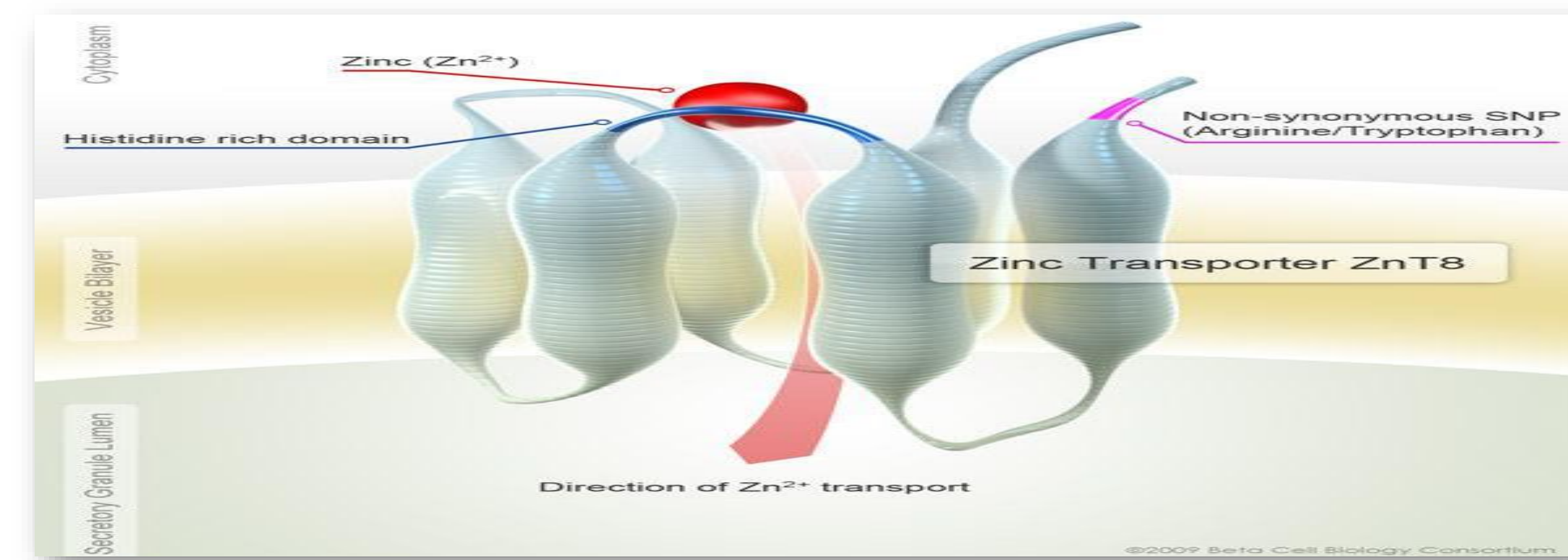
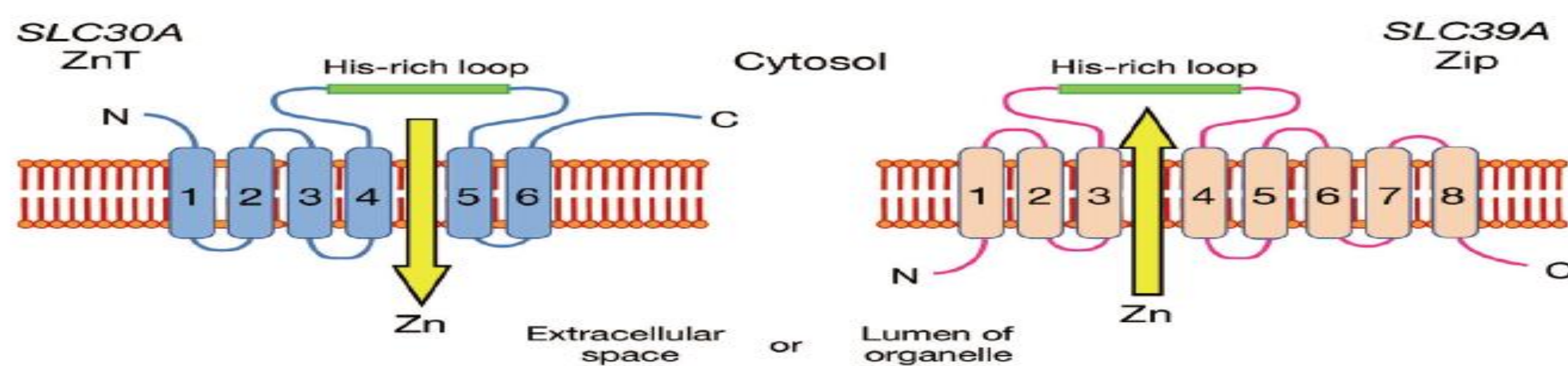
# Identification of zinc transporter ZnT8 in thyroid tissues from patients with immune and non-immune thyroid diseases

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## Objectives

### Zinc transporter - ZnT8 –function and localization



\* **Zn homeostasis** is regulated by ZnT (SLC30A gene family) and Zip (SLC39A gene family) zinc transporters.

\* **Zinc transporter 8** is localized in insulin containing secretory granule membrane and transports zinc from the cytosol into the vesicles. **ZnT8** contains six transmembrane domains and a histidine-rich loop between transmembrane domains 4 & 5, which is the putative zinc binding domain.

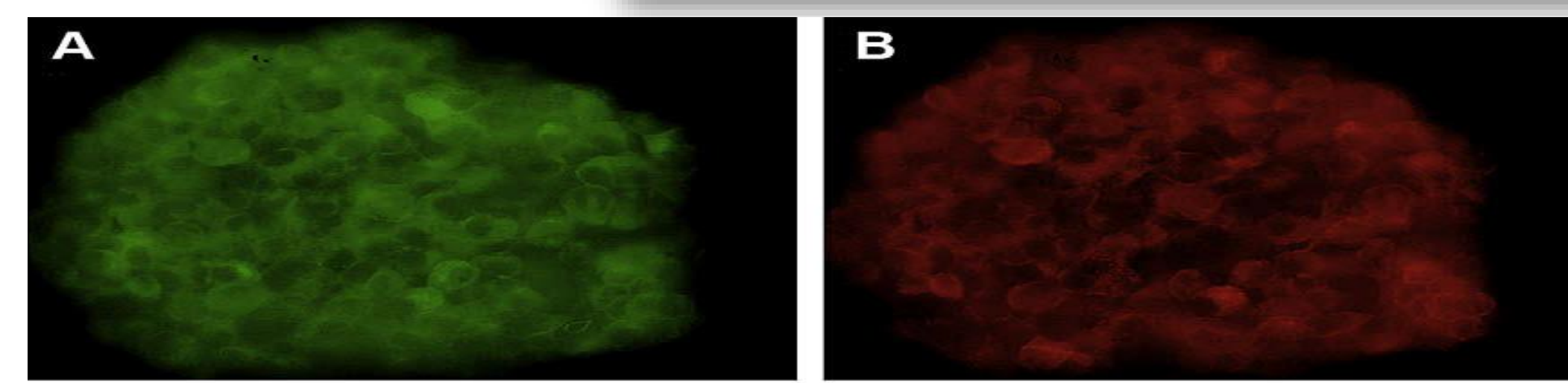
\* **Human SLC30A8 gene** is located to **chromosome 8** at the position **q24.11** which contains 8 exons and encoding a 369 amino acid ZnT8 protein.

\* **Zn** is important for the generation of proinflammatory cytokines: IL-6, TNF- $\alpha$  after LPS stimulations and development of T cells.

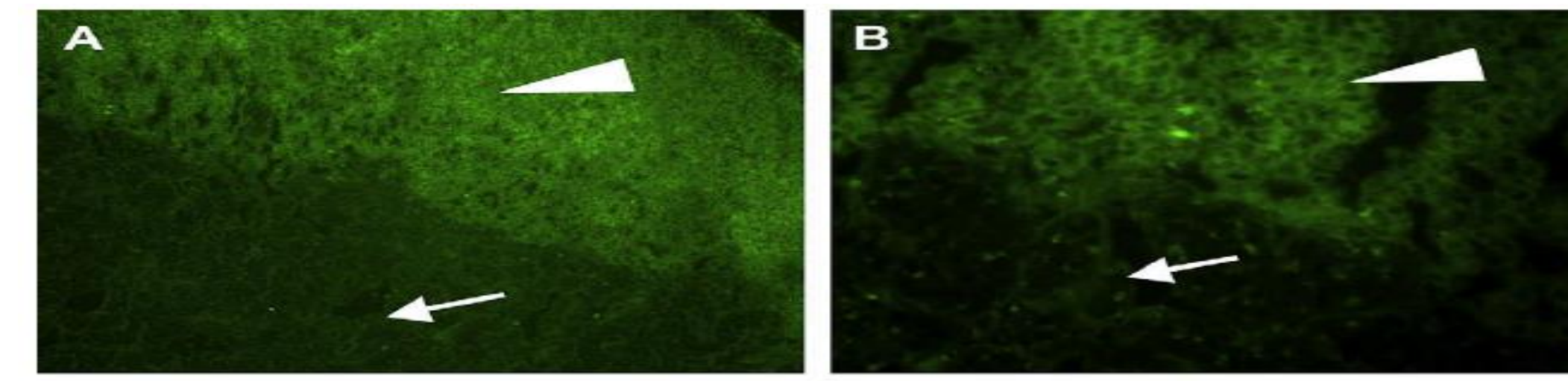
\* **Zinc deficiency** induces thymic atrophy, lymphopenia, suppression of cytolytic T cell responses & NK activity.

\* **Zinc protects** pancreatic B-cells from cytokine-induced destruction, which is observed in patients with DT1 & DT2.

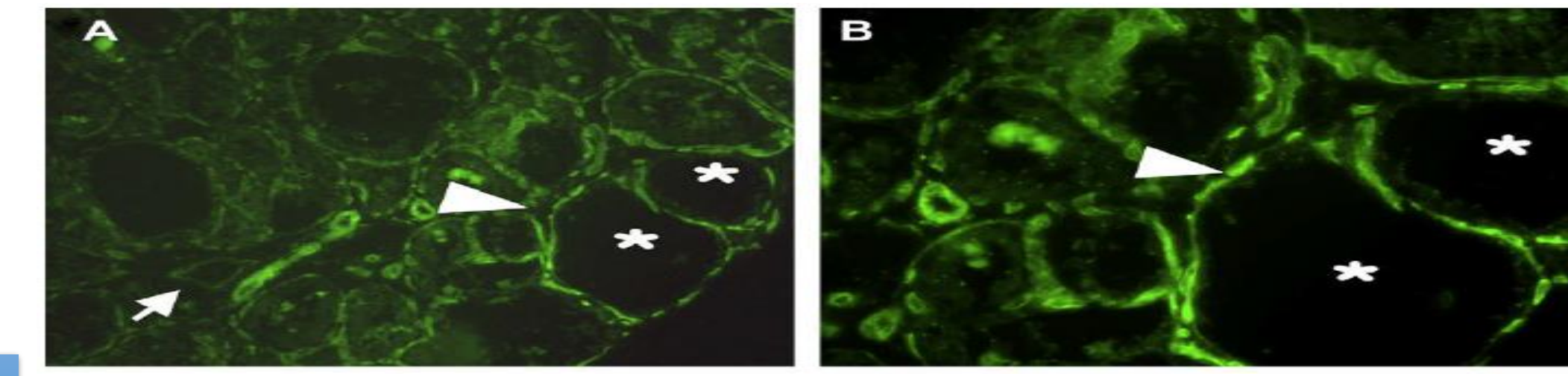
Kawasaki E. *Endocrine Journal* 2012, 59 (7), 531-537



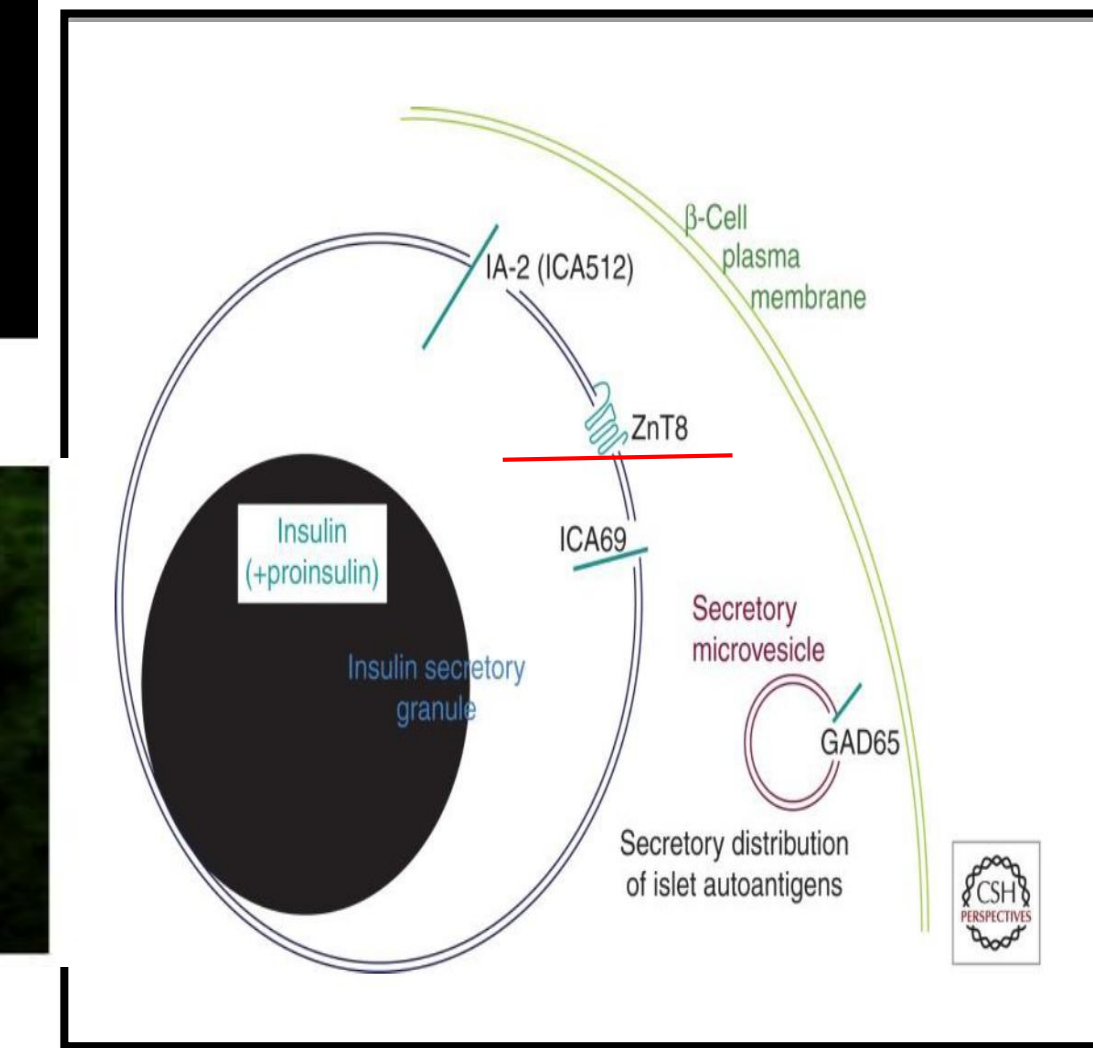
Localization of ZnT8 transporter in a pancreatic  $\beta$ -cells



Localization of ZnT8 transporter in adrenal cortex in animal model



Immunolocalization of ZnT8 transporter using anti-ZnT8 antibodies (a, b) in epithelium of thyroid follicular cells in animal model.



Murgia C et al. *Nutrition, Metab. & Cardiovascular Dis* (2009) 19, 431-439

## Material and Methods

We studied the expression of ZnT8 transporter in thyroid tissues from patients with immune and non-immune thyroid diseases. The study was performed in thyroid tissues after thyroidectomy from patients with thyroid nodular goiter (n=17, mean age 17.8 years  $\pm$  4) and cases with Graves' disease (n=20, mean age 16.6 years  $\pm$  2.4).

	Graves' disease	TNG	*p
Weight(kg)	54 $\pm$ 5.28	58.2 $\pm$ 7.8	p<0.01
Height (cm)	158.6 $\pm$ 4.3	156 $\pm$ 8	NS
fT4 (ng/dl)	1.9 $\pm$ 0.63	1.2 $\pm$ 0.46	p<0.01
fT3 (ng/dl)	5.78 $\pm$ 0.5	3.2 $\pm$ 0.38	p<0.01
TSH ( $\mu$ U/ml)	0.87 $\pm$ 2.37	3.04 $\pm$ 0.72	p<0.01
TRAb (U/I)	12.5 $\pm$ 0.31	0.4 $\pm$ 0.22	p<0.0001
anti-TGAb (IU/mL)	620.98 $\pm$ 240.34	98.6 $\pm$ 40.6	p<0.001
anti-TPOAb (IU/mL)	482.2 $\pm$ 62.43	56 $\pm$ 32.3	p<0.01
treatment	methimazole	L-thyroxine	

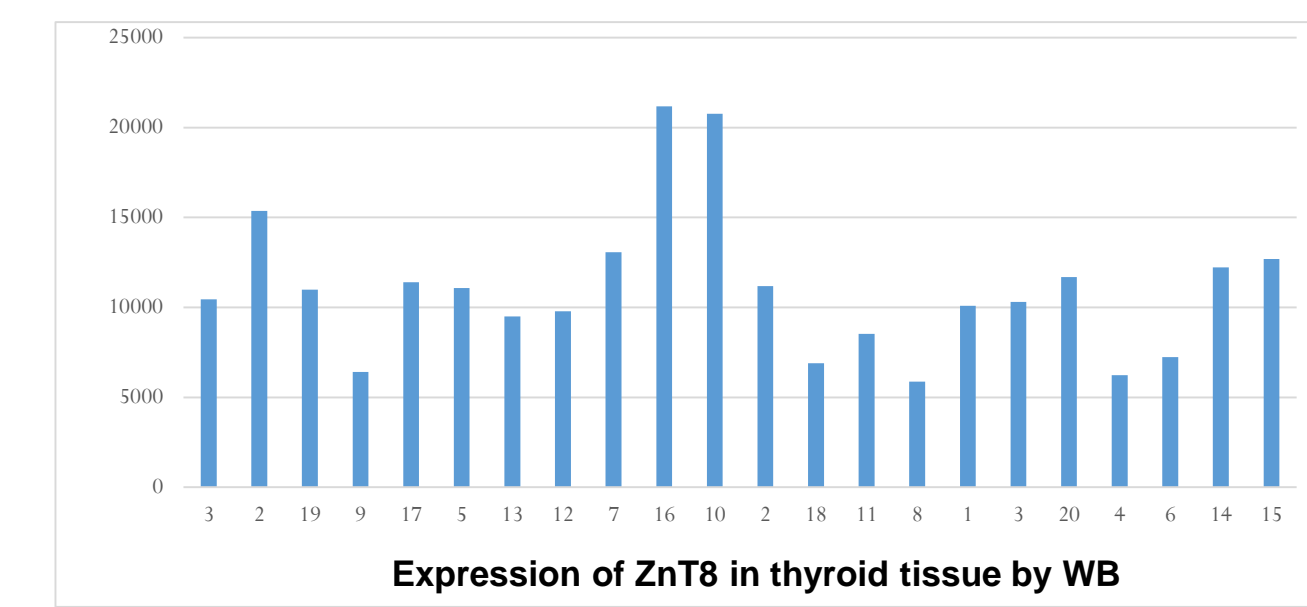
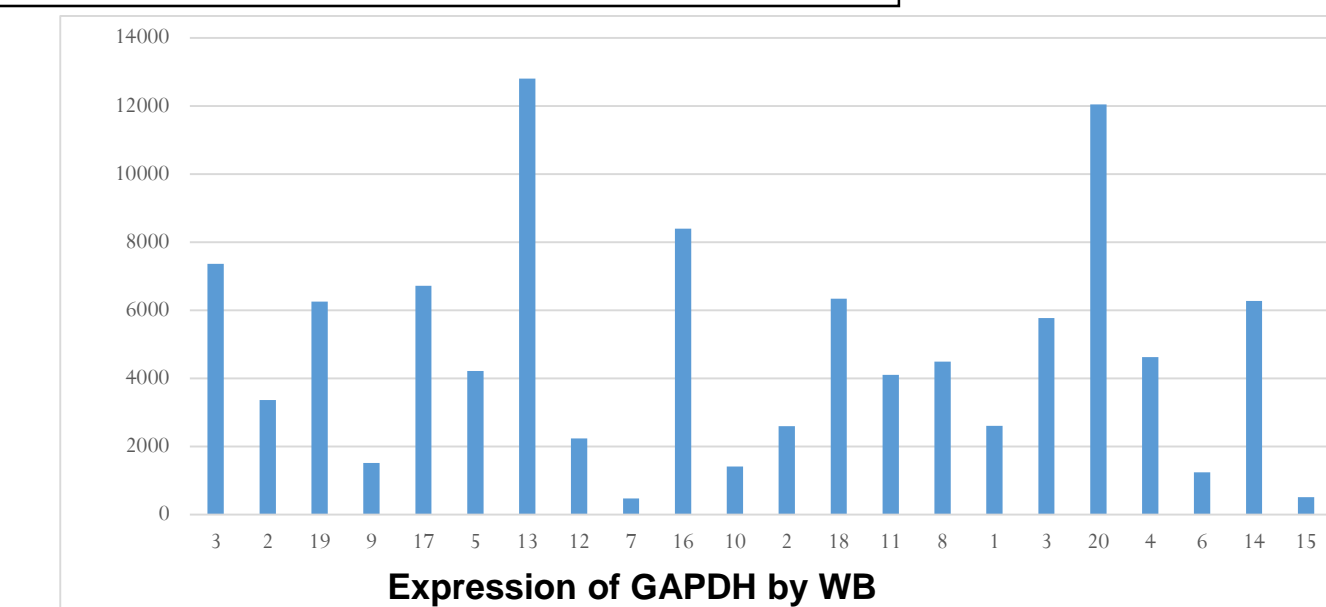
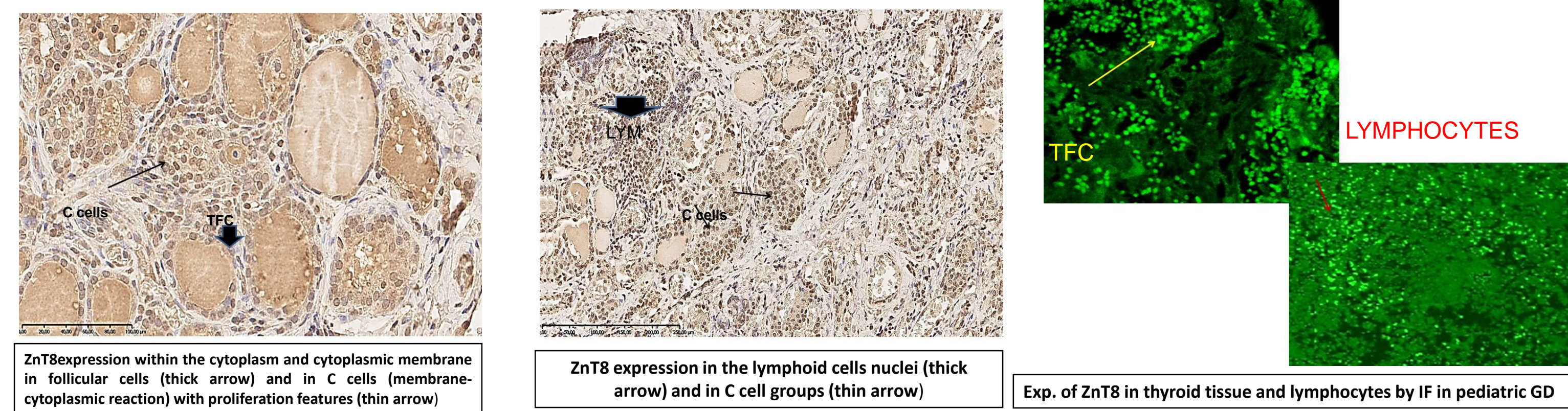
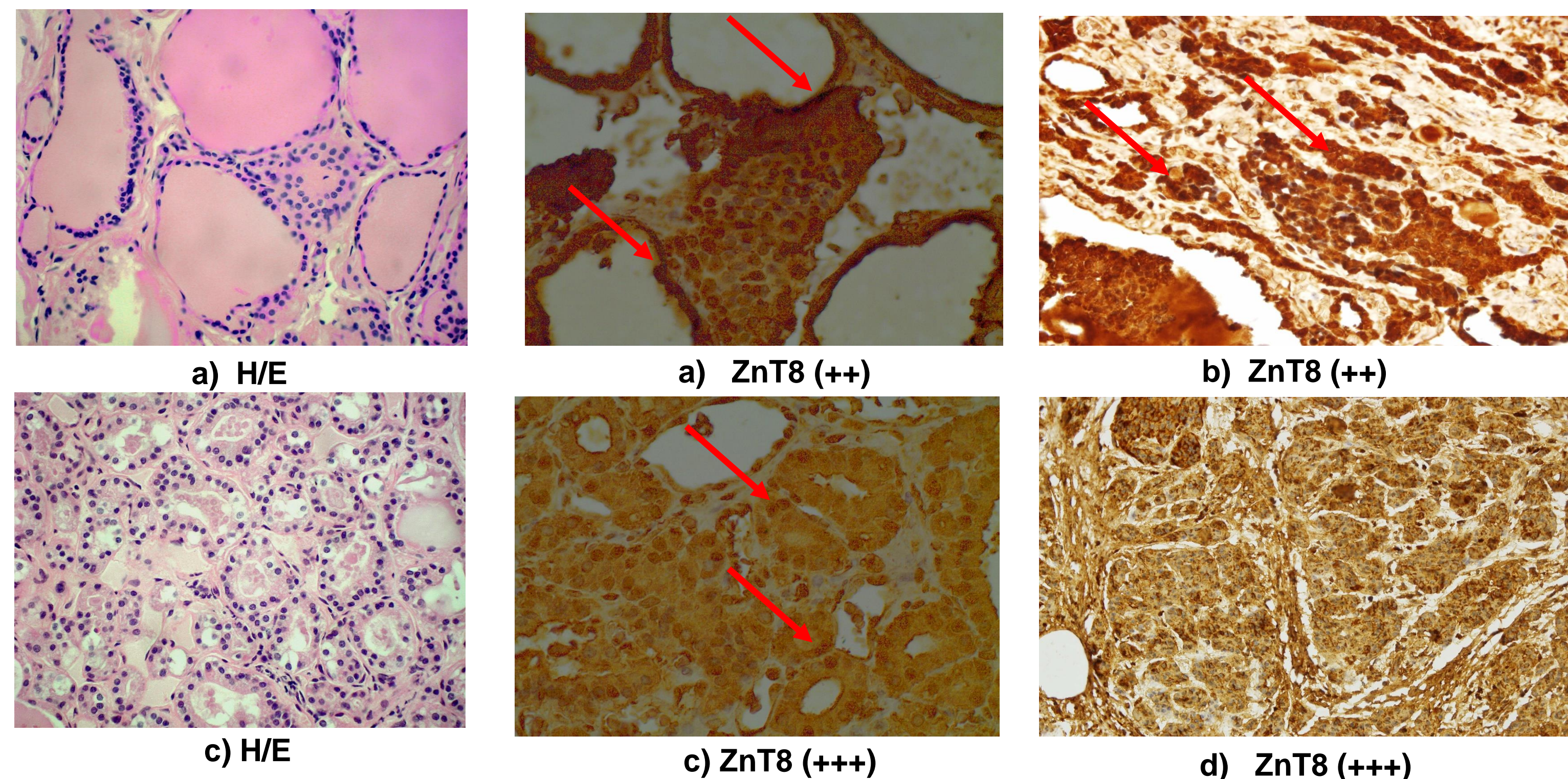
**Method:** The ZnT8 expression protein was evaluated using **immunohistochemistry**. The specimens were paraffin embedded tissues, derived from the pediatric patients, who had thyroid nodular goiter or GD. The antibody against ZnT8 was goat polyclonal antibody (Santa Cruz Biotechnology USA; sc-98243). The antigen retrieval was done using high pH (PTLink DAKO) and antibody was incubated in 4°C overnight in 1:50 dilution. The patients with pancreatitis were as a positive controls. The intensity and the proportion of stained cells were determined by examining the entire slide and section as:

+ (low staining intensity in less than 10% cells in the section); ++ (moderate staining intensity in 10-40% cells in the section); +++ (high and diffuse staining intensity in more than 50% cells in the section).

**Protein extraction and Western Blot:** The samples were homogenized in ice-cold RIPA (radioimmunoprecipitation assay) buffer containing protease and phosphatase inhibitors (Roche Diagnostics GmbH, Mannheim, Germany) for 1 min at 4°C. The homogenates were then incubated on the ice for 40 minutes. Next, homogenates were spin for 30 min at 10000 g at 4°C. After centrifugation the supernatant was collected and transferred into new eppendorf tubes. Total protein concentration was determined using BCA protein assay kit with BSA (bovine serum albumin) as a standard. Samples were boiled at 95°C for 10 minutes in sample buffer containing 2-mercaptoethanol. Protein (40  $\mu$ g) was subjected to 10% SDS-PAGE (sodium dodecyl sulfate-polyacrylamide gel electrophoresis) and transferred into nitrocellulose membranes. After blocking membranes in TTBS buffer (50 mM Tris-HCl, 130 mM NaCl and 0.05 % Tween-20) containing 5% nonfat dry milk for 90 min at room temperature membranes were incubated overnight at 4°C with the corresponding antibodies at a dilution of 1:500. Primary antibodies (Zn-T8 and GAPDH) were purchased from Santa Cruz Biotechnology. Thereafter the membranes were incubated with anti-rabbit IgG (for Zn-T8) and anti-mouse IgG (for GAPDH) horseradish peroxidase-conjugated secondary antibody (1:3000; Santa Cruz Biotechnology, USA). Immunoreactive protein bands were visualized by using an enhanced chemiluminescence substrate (Thermo Scientific, USA) and quantified by densitometry (Biorad, USA). Equal protein concentrations were loaded in each lane as confirmed by Ponceau staining on the blot membrane. Protein expression was normalized to GAPDH and expressed in arbitrary units.

## Results

**Identification of ZnT8 transporter in thyroid tissues by immunohistochemistry in patients with:** a) thyroid nodular goiter, b) C cells hyperplasia, c) Graves' disease & d) pancreatitis (positive control)



## Conclusions

1. Expression of ZnT8 transporter was identified in the thyroid tissues from paediatric patients with Graves' disease and nontoxic nodular goiter.
2. ZnT8 transporter expression was found both in thyroid follicular cells and C cells.
3. Predominant expression of ZnT8 in immune than in non-immune thyroid disorders may suggest potential role of ZnT8 as a new thyroid autoantigen but it requires further study on a larger cohort.