

Evaluation of molecular characteristics and steroid metabolomics in a large cohort P1-162 of children with 3^β-hydroxysteroid dehydrogenase 2 deficiency

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Context: Deficiency of 3β-hydroxysteroid dehydrogenase 2 (3βHSD2) causes a very rare form of congenital adrenal hyperplasia (CAH) known as 3^βHSD2 deficiency, which is a consequence of biallelic HSD3B2 gene defects. The estimated prevalence is less than 1/1,000,000 live births. Knowledge of comprehensive steroid metabolome patterns in 3βHSD2 deficiency is scarce. **Objective:** We aimed to investigate phenotypical, molecular, and biochemical characteristics, as well as the genotype-phenotype relationship in patients with 3βHSD2 deficiency. We evaluated steroid hormone profiles in individuals with homozygous and heterozygous HSD3B2 gene defects, mutation-negative "functional 3^βHSD2 deficiency", and patients with 21-hydroxylase deficiency (21-OHD).

Setting: Multi-centre, cross-sectional study in nine tertiary pediatric endocrinology clinics in

| ble 1. Sequence variations and | d genotype-phenotype | relationships in 31 | children with 3βHSD2 deficiency | |
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| Number of cases/families (n/n) | Genomic co-ordinates and nucleotide change (Genome assembly GRCh37.p13) | Protein (prediction) | cDNA position and nucleotide change (transcript NM_001166120, ENST00000543831) | PROVEAN | ariant Effect | PolyPhen2 (HumXar) | Mutation taster | Case report (DOI) | First functional characterization report (DOI) | Apparent activity in intact cells | Karyotype | Age(s) at diagnosis | Clinical type | Salt- wasting | DSD |
| 1/1 | g.4619_4620delCA chr1:119962172_119962173delC | p.H92Qfs*32 | c.274_275delCA cDNA.523_524delCA | ND | ND | ND | Disease causing | Present study | Present study | ND | 46, XX | Newborn | Classical | + | + |
| 1/1 | g.6891T>A chr1:119964444T>A | p.L107Q | c.320T>A cDNA.569T>A | -5.39 | 0.000 | 1 | Disease causing | Present study | Present study | ~12% of wild-type enzyme activity | 46, XY | Newborn | Classical | + (presentation and stress), normal potassium | + |
| 2/1 | g.7000_7001insAA chr1:119964553_119964554insA A | p.E144Kfs*31 | c.429_430insAA cDNA.678_679insAA | ND | ND | ND | Disease causing | Present study | Present study | ND | 46, XY (n=1) 46, XX (n=1) | Newborn-2 months | Classical | + | + (46, XY) - (46, XX) |
| 4/4 | g.7223T>C chr1:119964776T>C | p.S218P | c.652T>C cDNA.901T>C | -1.93 | 0.101 | 0.117 | Polymorphism | Present study | 10.1111/cen.12394 | <1-10% of wild-type enzyme activity [≇] | 46, XY (n=3) 46, XX (n=1) | Newborn-7 months | Classical | + (presentation and stress), normal potassium | + (46, XY) - (46, XX) |
| 2/1 | g.7304G>C chr1:119964857G>C | p.A245P | c.733G>C cDNA.982G>C | -1.57 | 0.196 | 0.979 | Polymorphism | Present study | 10.1210/mend.7.5.8316254 [§] 10.1210/jcem.84.12.6288 [©] | ~10% [§] and ~35% [©] of wild-type enzyme activity | 46, XY (n=2) | ~2 years | Classical | + (presentation and stress), normal potassium | + |
| 1/1 | g.7482T>C chr1:119965035T>C | p.L304P | c.911T>C cDNA.1160T>C | -5.66 | 0.009 | 0.783 | Disease causing | Present study | 10.1007/s11033-019- 04809-4 | ND | 46, XY | Newborn | Classical | + | + |
| 1/1 | g.7505delC chr1:119965058delC | p.F314Sfs*54 | c.934delC cDNA.1183delC | ND | ND | ND | Disease causing | Present study | Present study | ND | 46, XY | Newborn | Classical | + | + |
| 2/1 | g.7530_7531insC chr1:119965083_119965084insC | p.L321Ifs*4 | c. 959_960insC c.DNA.1208_1209ins C | ND | ND | ND | Disease causing | Present study | Present study | ND | 46, XY (n=1) 46, XX (n=1) | Newborn | Classical | + | + (46, XY) - (46, XX) |
| 14/11 | g.7538A>G chr1:119965091A>G | p.N323D | c.967A>G cDNA.1216A>G | -4.09 | 0.006 | 0.999 | Disease causing | Present study | 10.1007/s11033-019- 04809-4 | <5% of wild-type enzyme activity | 46, XY (n=7) 46, XX (n=7) | Newborn-8 months | Classical | + | + (46, XY) - (46, XX) |
| 2/1 | g.7634T>C chr1:119965187T>C | p.W355R | c.1063T>C cDNA.1312T>C | -12.43 | 0.000 | 0.996 | Disease causing | 10.4274/jcrpe.3306 | Present study | <5% of wild-type enzyme activity | 46, XY (n=2) | Newborn | Classical | + | + |
| 1/1 | g.7647T>C chr1:119965200T>C | р.L.359Р | c.1076T>C cDNA.1325T>C | -3.12 | 0.004 | 0.913 | Disease causing | Present study | Present study | <5% of wild-type enzyme activity | 46, XX (n=1) 46, XY (n=1) | 1-2 months | Classical | + | - |

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Patients or Other Participants: Children with homozygous 3βHSD2 deficiency (n=31), individuals with heterozygous 3βHSD2 deficiency (n=31), children with classical 21-OHD (n=57), functional 3βHSD2 deficiency (n=18), and healthy controls (n=172).

Main Outcome Measures: A structured questionnaire was used to assess clinical and biochemical phenotype data. Genetic analysis of HSD3B2 was performed using Sanger sequencing. We measured Δ 5-to- Δ 4 steroids and 11-oxygenated C19 and rogens in serum and urine by mass spectrometry. Novel HSD3B2 mutations were studied in silico and by in vitro enzyme kinetic assays (Fig 1).

Results: Eleven homozygous (6 novel) in 31 children from 24 families (19 male/12 female; mean age: 6.6 ± 5.1 yrs) were identified (Fig 2A). The missense variants >5% of wild-type 3βHSD2 activity in vitro were associated with non-salt losing clinical phenotype (Table 1, Fig **2B)**. There was a significant genotype-phenotype-steroid metabolome correlation in patients with 3 β HSD2 deficiency (Fig 3). The plasma ratio of (17OH-Pregnenolone + Pregnenolone + DHEA)/(170HProgesterone + Progesterone + Androstenedione + Cortisol) was superior to (17OH-Pregnenolone/Cortisol) to discriminate 3βHSD2 deficiency from the other groups. Heterozygote carriers and functional 3 β HSD2 deficiency patients showed higher Δ 5-to- Δ 4 steroids than controls (Fig 4A, 4B, 5A). 11-oxygenated androgens were significantly lower in patients with 3βHSD2 deficiency (Fig 5B).





Figure 4. Assessment of two baseline Δ5-to-Δ4 steroid ratios in the diagnosis and differential diagnosis of 3βHSD2 deficiency. Two baseline plasma adrenal Δ5-to-Δ4 adrenal steroid ratios were compared between the individuals with 3BHSD2 deficiency, 3BHSD2 deficiency-like conditions, heterozygous 3BHSD2 deficiency, 21-OHD and non-CAH control groups. Grey areas show control groups stratified according to age. Both of these ratios were very efficient to diagnose and to differentiate 3BHSD2 deficiency from the other groups.

Figure 2. Molecular characteristics of HSD3B2 mutations. (A) Schematic presentation of HSD3B2 with all known mutations and novel mutations detected in this study. Non-coding exonic segments are indicated as white boxes. Most of the previously reported mutations are located in exon four. Pathogenic missense/nonsense mutations are shown in the upper panel, while insertion/deletions are indicated in the lower panel. Novel mutations reported in this study are shown in red. Mutations with *in vitro* >5% residual 3βHSD2 activity are highlighted in green boxes. (B) The activities of wildtype and mutant HSD3B2 enzymes. The graph shows the relative activities of wildtype and mutant HSD3B2 expressed in COS-1 cells following incubating with pregnenolone for 1 h. The Western blot on the top shows the levels of HSD3B2 in the cells, and the numbers on top

According to our results obtained from this large cohort,

Conclusions:

There is a good correlation between glucocorticoid and mineralocorticoid functions with in vitro and biochemical enzyme activity in 3βHSD2 deficiency, whereas genital and gonadal phenotype and behaviour are more complex and variable.

of the protein bands are the relative protein levels after quantitation of the images.



Figure 3. The effect of the severity of HSD3B2 mutation and karyotype on 3βHSD2 enzyme activity index. The effect of the severity of HSD3B2 mutation (A) and karyotype (B) on 3 β HSD2 enzyme activity index that is represented by simultaneously measured Δ 5 (pregnenolone, 17 α hydroxypregnenolone, DHEA) to $\Delta 4$ (progesterone, 17α -hydroxyprogesterone, and rostenedione and cortisol) steroids ratio. *HSD3B2* mutations causing a frameshift of gene sequence (p.H92Qfs*32, p.E144Kfs*31, p.F314Sfs*54, p.L321Ifs*4) result in a severely impaired 3βHSD2 enzyme activity index compared to missense mutations (p.L107Q, p.S218P, p.A245P, p.L304P, p.N323D, p.W355R, p.L359P). The difference of this ratio in patients with mild missense (p.L107Q, p.S218P, p.A245P; HSD3B2 activity ~5-10% of wild type) and severe missense (p.L304P, p.N323D, p.W355R and p.L359P; HSD3B2 activity <5% of wild type) mutations was not statistically significant (p=0.62) (A). 3βHSD2 enzyme activity index was similar in 7 girls and 7 boys with classical 3βHSD2 deficiency due to homozygous p.N323D mutation (p=0.62) (B).

In contrast to common knowledge, mineralocorticoid deficiency is not apparent in 1/3 of the cases.

- This 46, XY DSD is a "sine qua non" in affected males whereas ambiguous genitalia is only rarely seen in affected 46, XX individuals due to decreased production of potent androgens via classical or alternative pathways.
- On the other hand, premature pubarche is very common on either sex in 3βHSD2 deficiency.
- Spared mineralocorticoid function and unvirilized genitalia in females may lead to misdiagnosis and underestimation of the frequency of 3βHSD2 deficiency.
- Mass spectrometry-based measurements of $\Delta 5$ -to- $\Delta 4$ steroids are very sensitive and specific to diagnose and differentiate 3^βHSD2 deficiency from clinically look-alike conditions.
- Heterozygous 38HSD2 deficiency impairs biochemical 38HSD2 enzyme activity but does not cause a clinically significant phenotype.
- The term "non-classic or late-onset" form of 3BHSD2 deficiency, if it is existent, should only be used following genetic confirmation. The role of the regulators of 3BHSD2 in the pathogenesis of adrenal androgen excess needs to be elucidated.
- The correct diagnosis of 3BHSD2 deficiency is not only essential for the proper clinical management in infancy and childhood but also for the surveillance of gonadal functions and fertility of the patients in later life.





